



# Current Understanding of the Correlation of Lignin Structure with Biomass Recalcitrance

Mi Li<sup>1</sup>, Yunqiao Pu<sup>1</sup> and Arthur J. Ragauskas<sup>1,2,3\*</sup>

<sup>1</sup> BioEnergy Science Center, Biosciences Division, Joint Institute of Biological Science, Oak Ridge National Laboratory, Oak Ridge, TN, USA, <sup>2</sup> Department of Chemical and Biomolecular Engineering, University of Tennessee Knoxville, Knoxville, TN, USA, <sup>3</sup> Department of Forestry, Wildlife, and Fisheries, Center for Renewable Carbon, University Tennessee Institute of Agriculture, Knoxville, TN, USA

## OPEN ACCESS

### Edited by:

John D. Wade,  
Florey Institute of Neuroscience and  
Mental Health, Australia

### Reviewed by:

Irina Bakunina,  
Pacific Institute of Bioorganic  
Chemistry RAS, Russia  
Alexander Shekhtman,  
University at Albany, SUNY, USA

### \*Correspondence:

Arthur J. Ragauskas  
aragausk@utk.edu

### Specialty section:

This article was submitted to  
Chemical Biology,  
a section of the journal  
Frontiers in Chemistry

**Received:** 16 August 2016

**Accepted:** 02 November 2016

**Published:** 18 November 2016

### Citation:

Li M, Pu Y and Ragauskas AJ (2016)  
Current Understanding of the  
Correlation of Lignin Structure with  
Biomass Recalcitrance.  
Front. Chem. 4:45.  
doi: 10.3389/fchem.2016.00045

Lignin, a complex aromatic polymer in terrestrial plants, contributes significantly to biomass recalcitrance to microbial and/or enzymatic deconstruction. To reduce biomass recalcitrance, substantial endeavors have been exerted on pretreatment and lignin engineering in the past few decades. Lignin removal and/or alteration of lignin structure have been shown to result in reduced biomass recalcitrance with improved cell wall digestibility. While high lignin content is usually a barrier to a cost-efficient application of bioresources to biofuels, the direct correlation of lignin structure and its concomitant properties with biomass remains unclear due to the complexity of cell wall and lignin structure. Advancement in application of biorefinery to production of biofuels, chemicals, and bio-derived materials necessitates a fundamental understanding of the relationship of lignin structure and biomass recalcitrance. In this mini-review, we focus on recent investigations on the influence of lignin chemical properties on bioprocessability—pretreatment and enzymatic hydrolysis of biomass. Specifically, lignin-enzyme interactions and the effects of lignin compositional units, hydroxycinnamates, and lignin functional groups on biomass recalcitrance have been highlighted, which will be useful not only in addressing biomass recalcitrance but also in deploying renewable lignocelluloses efficiently.

**Keywords:** biomass recalcitrance, lignin structure, cell wall, pretreatment, enzymatic hydrolysis

## INTRODUCTION

The conversion of renewable lignocellulosic biomass to fuels and valuable co-products, usually referred as biorefining, has been advanced recently (Ragauskas et al., 2006). However, transition from fossil-based to biomass-based products using current feedstocks and technologies has been challenged by the inherent resistance of plant cell walls to microbial and enzymatic deconstruction, namely, recalcitrance (Himmel et al., 2007). While other factors could not be neglected, the presence of lignin (ca. 15–35% by weight) is one of the most significant recalcitrance contributors, which escalates the processing costs (i.e., necessitated pretreatment and enlarged enzyme amount; Mosier et al., 2005; Pu et al., 2011, 2013). Lignin is an amorphous and complex aromatic polymer providing terrestrial plants mechanical support, stress response, pathogen resistance, and water transport (Boerjan et al., 2003). By bonding or embedding with other biopolymers (cellulose and hemicellulose), lignin strengthens the integrity and rigidity of the plant cell wall yielding a complex macro-molecular assembly (**Figure 1**). This lignin-polysaccharides matrix renders cell

walls recalcitrance for biorefining. Depending on biological species, lignin in plants derives primarily from three phenylpropanoid monolignols—*p*-coumaryl, coniferyl, and sinapyl alcohols (Li M. et al., 2016; Yoo et al., 2016b), which give rise to *p*-hydroxyphenyl (H), guaicyl (G), and syringyl (S) units, respectively (Figure 1). The inter-linkages between these subunits are  $\beta$ -O-4',  $\beta$ -5',  $\alpha$ -O-4', 4-O-5',  $\beta$ - $\beta'$  in common, and  $\beta$ -1', and 5-5' in minor amount.

In biomass, lignin content, structure, subunits, and linkages with polysaccharides own their remarkable importance to cell wall recalcitrance, usually gauged by biomass enzymatic hydrolyzability (Zeng et al., 2014; McCann and Carpita, 2015). Lignin limits the enzymatic hydrolysis of biomass through two primary mechanisms: restricting polysaccharides accessibility (physical barrier and/or lignin-carbohydrate complexes) and non-productive binding with enzymes (inhibition).

Current strategies to reduce lignin involved biomass recalcitrance mainly include (i) pretreatment technologies (Mood et al., 2013; Pu et al., 2013), (ii) perturbing lignin biosynthesis (Bonawitz and Chapple, 2010; Simmons et al., 2010), and (iii) enzyme engineering and modification (Güven et al., 2010). Whichever strategy is used, fundamental understanding of the influence of lignin's physicochemical properties on biomass recalcitrance is crucial to advance the biologically-based biorefinery. This mini-review aims at recent findings on the relationship between lignin structure and biomass recalcitrance.

## LIGNIN DURING PRETREATMENT

Lignin restricts enzymatic hydrolysis commonly through physical barrier and non-productive/non-specific binding to enzymes. While many pretreatments (e.g., alkaline/organosolv) substantially remove lignin yielding biomass with enhanced cellulose accessibility, some thermochemical pretreatments such as HWP, DAP, SEP, and AFEX, usually with limited delignification, lead to reduced biomass recalcitrance too (Mood et al., 2013; Pu et al., 2015). The cell wall matrix disruption could be facilitated by cleavage of aryl ether linkages in lignin and LCC linkages, which increases pore volumes and cellulose accessibility. For example,  $\beta$ -O-4' linkages are significantly cleaved in HWP poplar (Samuel et al., 2013) and DAP switchgrass (Samuel et al., 2010). AFEX reduced lignin-hemicellulose ester linkages revealed by a substantial loss of ferulate groups in the residual lignin (Chundawat et al., 2011). In addition, the translocation and redistribution of lignin coalescing into lignin droplets of various morphologies have been found in corn stem and switchgrass during DAP and HWP, which related to increased cell wall accessibility (Donohoe et al., 2008; Pingali et al., 2010). It was found that the lignin-hemicellulose complex underwent phase separation during pretreatment

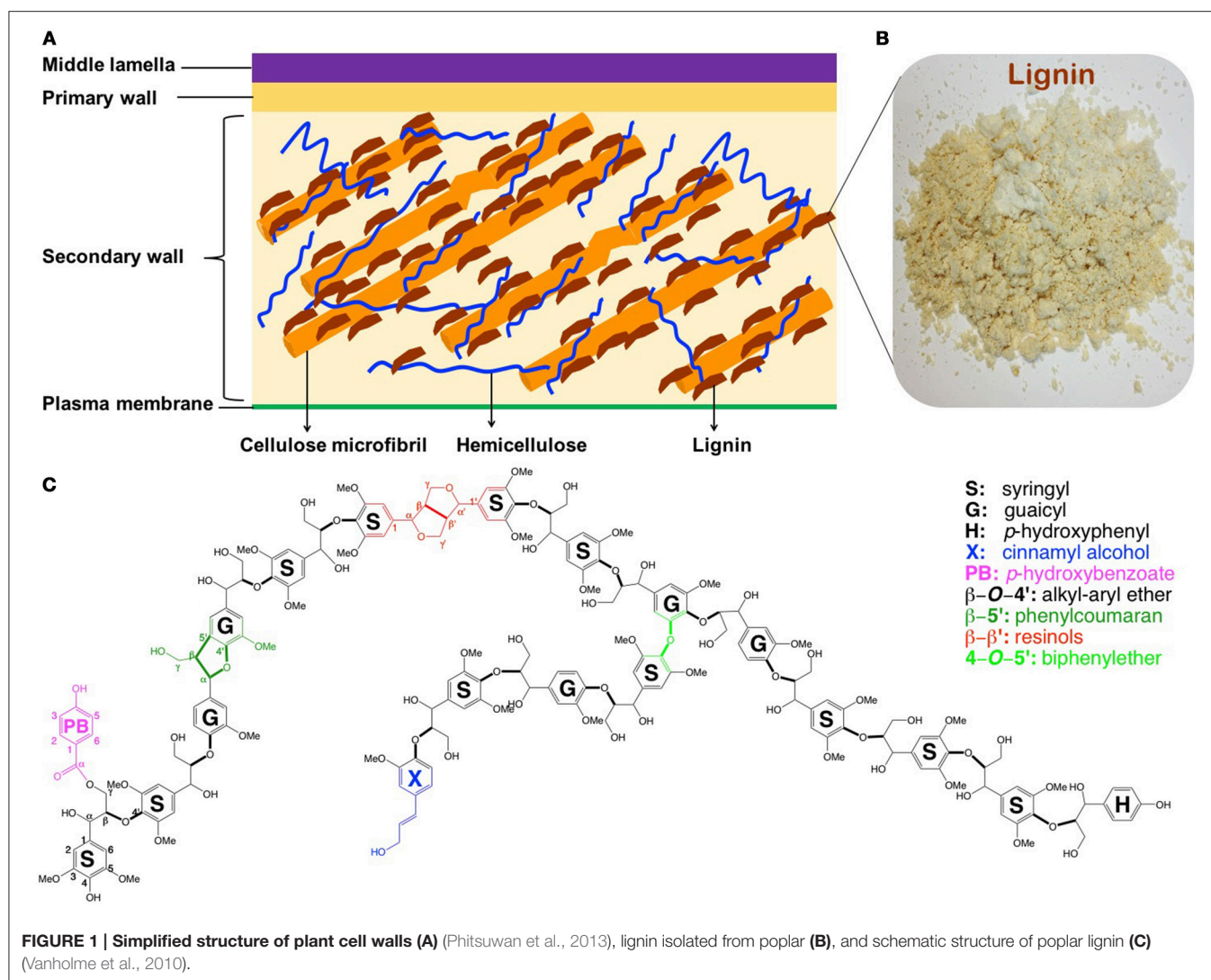
which drove the lignin aggregation followed by collapse, leading to increasing cell wall porosity (Langan et al., 2014). Using mixed poplar and Avicel, the lignin droplets from poplar was redeposited on the Avicel during HWP, causing 30–40% decrease of initial hydrolysis compared with Avicel control (Li H. et al., 2014). Interestingly, lignin-like droplets, pseudo-lignin, which inhibit enzymatic hydrolysis, can form from carbohydrates during acidic pretreatments (Sannigrahi et al., 2011; Hu et al., 2012).

## LIGNIN-ENZYME INTERACTIONS

Lignin has remarkable influence on enzymatic performance owing to lignin-enzyme interactions. Stemming primarily from hydrophobic, ionic, and hydrogen bond interactions with protein, lignin-enzyme interactions are highly influenced by the physicochemical properties of lignin (Nakagame et al., 2011a). Lignin is generally found inhibitory to cellulose hydrolyzability due to non-productive/non-specific binding with enzymes. For example, when added into the enzymatic hydrolysis of Avicel or pretreated biomass, several commercial lignins and isolated lignins from variable biomass origins apparently decreased saccharification (Nakagame et al., 2010, 2011c; Kim, 2012; Ko et al., 2015). However, cellulolytic lignin residues from corn stover and wheat straw had minimal impact on the hydrolyzability of pretreated biomass (Barsberg et al., 2013). Surprisingly, recent studies reported certain technical lignins with promoting effects, e.g., lignosulfonate (Wang et al., 2013; Zhou et al., 2013), organosolv lignin (Lai et al., 2014), Kraft lignin (Wang et al., 2015), and alkali lignin (Li Y. et al., 2016). These authors attributed the positive effect of lignin to the alleviation of non-productive binding *via* lignosulfonate-enzyme complex or surfactant protection. Several studies also suggested that lignins isolated from herbaceous plants had relatively less inhibition than woody biomass, likely because (i) branched lignin (G-lignin) is more inhibitory than linear lignin (S-lignin) and (ii) the formation of metal ion (e.g.,  $\text{Ca}^{2+}$ )-lignin complex could reduce lignin-enzyme interactions (Liu et al., 2010; Barsberg et al., 2013). In addition, the inhibitory effects of lignin depend on pretreatment severity. Lignin derived from more severely pretreated biomass exhibited more pronounced inhibition to the hydrolysis of Avicel because increased pretreatment severity resulted in more condensed structure (Nakagame et al., 2011c; Ko et al., 2015). Lignin repolymerization with increased C-C condensed structure presumably *via* the formation of carbonium ions can occur during HWP, DAP, and SEP (Pu et al., 2015). Lignin condensation associated with hydrophobicity influences lignin-enzyme interactions significantly. A few studies have shown that lignin with increased condensation from pretreated biomass tended to adsorb more enzymes, resulting in more inhibitory to cellulose hydrolysis (Yu et al., 2014; Ko et al., 2015; Huang et al., 2016; Yang and Pan, 2016).

Isolated lignin seems be more inhibitory to the hydrolysis of pure cellulose than lignocellulosic materials. Isolated Douglas-fir lignin decreased the hydrolysis yields of Avicel and

**Abbreviations:** AFEX, ammonia fiber expansion; 4CL: 4-coumarate:coenzyme A ligase; C3'H, *p*-coumaroyl quinate/shikimate 3'-hydroxylase; C4H, cinnamate 4-hydroxylase; CAD, cinnamyl alcohol dehydrogenase; CCR, cinnamoyl-coenzyme A reductase; CCoAOMT, caffeoyl-CoA O-methyltransferase; COMT, caffeic acid O-methyltransferase; DAP, dilute acid pretreatment; HCT, hydroxycinnamoyl CoA: shikimate hydroxycinnamoyl transferase; HWP, hot water pretreatment; SEP, steam explosion pretreatment.



pretreated softwood by 46 and 9%, respectively (Kumar et al., 2012). Kraft lignin and lignosulfonate inhibited pure cellulose saccharification but enhanced the hydrolysis of pretreated biomass (Liu et al., 2010; Kim, 2012; Zhou et al., 2013; Wang et al., 2015). In comparison to pure cellulose, the complexity of lignocellulosic substrates probably plays a role in the lignin-enzyme interactions. As noted by Zhou et al., lignosulfonate interacts with both the bound and soluble lignin of the substrate (Zhou et al., 2013). Hydrolysis of the same biomass with different degree of sulfonation demonstrated different enhanced digestibility when sulfonated lignin was added (Wang et al., 2015). Another important finding was that non-productive/non-specific binding predominated for less accessible biomass; with increased cellulose accessibility of lignocellulose, the inhibitory effects of lignin dwarfed (Kumar et al., 2012). Therefore, the lignin-enzyme interactions, conventionally termed as “detrimental effect,” varies significantly on lignin chemistry, type of substrate as well as pretreatment techniques employed.

## LIGNIN MONOLIGNOL COMPOSITIONAL UNITS

The monolignol compositional units (relative abundance of H, S, and G) of lignin have been documented to affect biomass digestibility. Without pretreatment, several studies found that S/G ratio was negatively related to the enzymatic hydrolysis of untreated biomass, e.g., engineered poplar, eucalyptus mutants, and maize cell wall (Zhang et al., 2011; Papa et al., 2012; Min et al., 2013). The authors deduced the negative effect of S/G likely to a more efficient coverage of S-lignin (extended shape) than G-lignin (branching) on cellulose fibrils according to a proposed molecular model (Besombes and Mazeau, 2005a,b). However, a few other studies reported that the hydrolyzability of untreated biomass was not affected by S/G ratio, such as *Populus* natural variants with different S/G ratio (1.0–3.0) (Studer et al., 2011), high G (95%) and high S (91%) contained *Arabidopsis* (Li et al., 2010), and transgenic poplar lines with 87 and 93% S (Mansfield et al., 2012) showing basically similar hydrolysis efficiency vs.



their corresponding controls. It seems that the small hydrolysis improvement in lower S/G plants without pretreatment might be influenced by factors arising from other cell wall components, varied lignin content, or type of enzyme.

By contrast, S/G ratio in lignin has shown remarkable influence in pretreatment and the hydrolyzability of pretreated biomass. Enhanced saccharification upon S/G ratio has been reported on *Populus* (Studer et al., 2011) and *Arabidopsis* (Li et al., 2010) by HWP, and poplar by SEP (Mansfield et al., 2012). S-unit was preferentially removed relative to G-unit in poplar during flowthrough pretreatment (Trajano et al., 2013). In addition, S/G ratio is positively related with delignification of hardwood by green liquor and Kraft pulping (Santos et al., 2011; Min et al., 2013). According to these authors, high S/G ratio benefits pretreatment mainly due to (i) lignin depolymerization: S-lignin features higher level of labile  $\beta$ -O-4' linkages which are readily cleavable during pretreatment; and (ii) delignification: S-lignin with relatively higher occurrence of  $\beta$ - $\beta'$  bonds leads to lower molecular weight which could facilitate lignin migration and removal. However, inconsistent results have also been reported on the correlation of S/G ratio with saccharification efficiency, e.g., high S/G ratio was negatively associated with the saccharification of both NaOH and H<sub>2</sub>SO<sub>4</sub> pretreated *Miscanthus* (Xu et al., 2012; Li M. et al., 2014) and green liquor pretreated wheat straw (Xu et al., 2012; Li M. et al., 2014; Jiang et al., 2016). An unclear correlation has also been reported, such as green liquor and Kraft pretreated *Eucalyptus* (Papa et al., 2012; Santos et al., 2012), NaOH and H<sub>2</sub>SO<sub>4</sub> pretreated wheat and rice samples (Wu et al., 2013), and mild acid treated maize cell wall (Zhang et al., 2011). When the S/G ratio of the remaining lignin in pretreated biomass is studied, contradictory results also exist: a lesser proportion of non-condensed lignin in the pretreated biomass was reported to be beneficial to biomass hydrolysis as non-condensed lignin (high  $\beta$ -O-4') tended to be linear shape, likely a higher coverage over cellulose fibers (Zhang et al., 2011; Yeh et al., 2014); by contrast, others found high S/G ratio of lignin in the pretreated biomass was positively correlated with enzymatic digestibility as branched G-lignin gave rise to more physical barrier (Santos et al., 2012; Yu et al., 2014). These inconsistent results suggest that S/G ratio contributes only partially to biomass recalcitrance, and the complexities of biomass species and pretreatment with concomitant other cell wall structure changes likely shelter the effects arising from lignin composition.

Due to the low occurrence of H-lignin reported in natural plants, the influence of H-lignin on biomass recalcitrance is relatively less investigated. Interestingly, a few recent studies revealed that high H-lignin contained plants exhibited reduced biomass recalcitrance. For instance, transgenic Alfalfa (50–76% H) had increased alkaline extractability of lignin-like content vs. control (5% H) (Ziebell et al., 2010); the H/G ratio measured in the KOH-extractable lignin of wheat and rice had a strong positive correlation with glucose yield of pretreated biomass (Wu et al., 2013); H-rich (89%) *Arabidopsis* mutant exhibited significantly higher sugar yield than G-rich (96%) and S-rich (92%) mutants without pretreatment (Shi et al., 2016); hexose yield increased upon H level in alkali pretreated *Miscanthus* (Li

M. et al., 2014). The speculated reasons for the positive effect of H-lignin are (i) reduced lignin molecular weight, (ii) reduced cellulose crystallinity via H unit-glucan bonding; and (iii) higher linkage activities between H monomer than G and S.

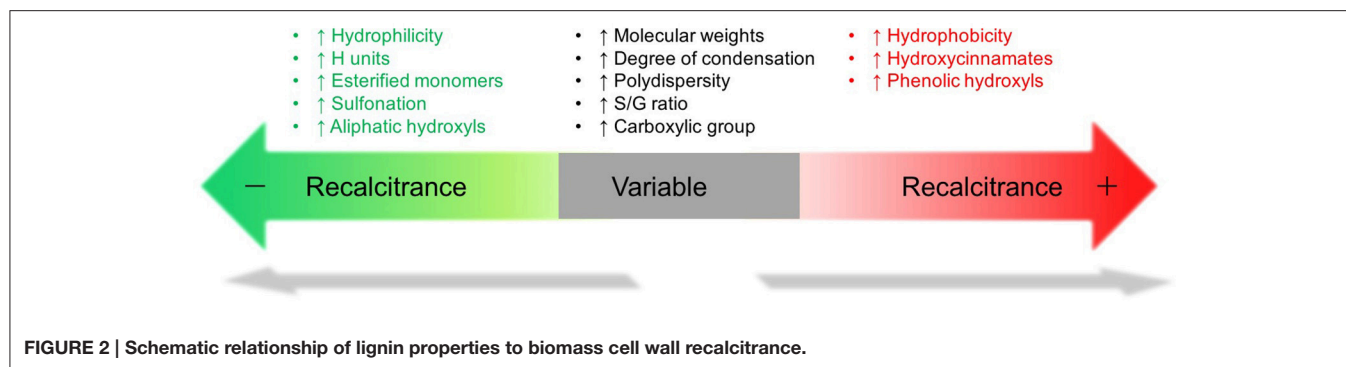
## HYDROXYCINNAMATES IN LIGNIN

Biomass recalcitrance is also related to the presence of hydroxycinnamates in lignin, mainly ferulates (FA) and *p*-coumarates (*p*CA) which are two important hydroxycinnamates in grass (Buranov and Mazza, 2008; Ralph, 2010). FA is reported to mediate the cross-linking of polysaccharides-polysaccharides and polysaccharides-lignin in cell wall (de O Buanafina, 2009; Azarpira et al., 2011). The presence of this cross-linking draws lignin to polysaccharides proximately resulting in an increased recalcitrance. Reduced FA-mediated cross-linking of lignin-polysaccharides in maize (Jung and Phillips, 2010) and silage (Jung et al., 2011) had improved digestibility. Due to the strong negative correlation of etherified phenols, FA content in forage crop silages was good predictor for *in vivo* cell wall digestibility (Taboada et al., 2010). To enhance biomass digestibility, one of the effects of pretreatment is to cleave FA cross-linkages and promote lignin degradation and coalescence (Li et al., 2012; Qin et al., 2015; Martínez et al., 2016; Yoo et al., 2016a). Rather than cross-linking inter-components in cell wall, *p*CA are usually esters pendantly linked on lignin ( $\gamma$ -position of S unit), but they are also found to acylate to polysaccharides (Petrik et al., 2014). The accumulation of *p*CA indicating lignin deposition level in plants is likely one reason for recalcitrance (Taboada et al., 2010). The *p*CA content of alkaline hydrogen peroxide pretreated grasses, proportional to lignin content, was negatively related with enzymatic digestibility (Li et al., 2012). A study of genetically modified maize lines with similar lignin level (12–15%) demonstrated that the *in vivo* cell wall digestibility is also negatively correlated with the esterified *p*CA and lignin content (Zhang et al., 2011). Due to the acylation with hemicellulose (arabinoxylan), removal of *p*CA together with xylan in sugar cane bagasse increased cellulose accessibility thereof saccharification efficiency (Martínez et al., 2016).

On the other hand, hydroxycinnamates bearing readily cleavable ester linkages was incorporated into lignification to reduce biomass recalcitrance (Ralph, 2010). Incorporation of hydroxycinnamates conjugate—rosmarinic acid, with monolignols into maize cell walls via artificial lignification had remarkably enhanced alkali extractability and digestibility of cell walls (Tobimatsu et al., 2012). More recently, by introducing monolignol ferulate into lignin monomer pool, the engineered poplar demonstrated an improved cell wall digestibility after alkaline pretreatment (Wilkerson et al., 2014).

## LIGNIN HYDROXYL AND CARBOXYLIC GROUPS

As a major driving force in lignin-enzyme interactions, the hydrophobicity of lignin changes upon the content of



hydroxyl and carboxylic groups, which has important influence on enzymatic hydrolysis. Generally, lignin hydrophobicity increases with phenolic hydroxyl (Ar-OH) and decreases with aliphatic OH content. High Ar-OH and low aliphatic OH amounts in lignin were related with higher cellulases adsorption affinity and binding strength (Yu et al., 2014; Huang et al., 2016). Lignin with increased Ar-OH had higher enzyme affinity thereof more inhibition to the hydrolysis of Avicel (Rahikainen et al., 2013; Guo et al., 2014). Yang and Pan recently found that Ar-OH (3.5–4.4 mmol/g lignin) blocked by hydroxypropylation had negligible effect on enzyme adsorption, however, reduced lignin inhibition significantly, which suggested Ar-OH was more related with other interaction for inhibitory effect (Yang and Pan, 2016). Carboxylic groups (COOH) in lignin alter the physicochemical effects of lignin by (i) increasing hydrophilicity; (ii) creating electrostatic charge; and (iii) enhancing hydrogen bond, which could impact lignin-enzyme interactions differently (Nakagame et al., 2011a). Increased COOH in lignin enhanced the hydrolysis efficiency of Avicel as COOH may alleviate the non-productive binding by increasing hydrophilicity and electrostatic repulsion force of lignin to enzymes (Nakagame et al., 2011b; Moxley et al., 2012). Increasing the hydrophilicity of lignin via carboxylation and sulfonation reduced inhibitory effect substantially (Yang and Pan, 2016). However, the correlation of Ar-OH and COOH with enzymatic hydrolysis becomes complicated when different botanical origins and pretreatment methods are used due to the complexity of substrates. For example, high Ar-OH was related with increased sugar yield of Kraft pretreated hardwood and acid pretreated corn (Moxley et al., 2012; Santos et al., 2012), while inconsistent correlations between COOH and enzyme adsorption have been reported on lignins from different botanical origins (Rahikainen et al., 2013; Guo et al., 2014; Yu et al., 2014).

## OTHER LIGNIN ASSOCIATED FACTORS

Other factors of lignin such as molecular weight and polydispersity were found associated with biomass recalcitrance but the correlation is inconclusive (Figure 2). The molecular weight of alkali lignin and liginosulfonate had an opposite effect on enzymatic hydrolysis (Zhou et al., 2013; Li Y. et al., 2016) and

no clear impact of the molecular weights on enzyme adsorption was found (Pareek et al., 2013; Guo et al., 2014). Although earlier study indicated that lignin with lower polydispersity thereby higher plasticity, interacts favorably with proteins (Berlin et al., 2006), a few recent reports had difficulty to correlate lignin polydispersity with saccharification/enzyme adsorption (Pareek et al., 2013; Guo et al., 2014; Yang and Pan, 2016).

An alternative approach to enhance saccharification is to manipulate lignin biosynthesis pathway in plant for reduced recalcitrance (Phitsuwan et al., 2013). A few strategies used for lignin biosynthesis perturbation are (i) decreasing lignin content; (ii) relocating lignin deposition; (iii) altering lignin subunits; and (iv) modification of lignin backbone and linkages with carbohydrates (Bonawitz and Chapple, 2010; Simmons et al., 2010; Cesarino et al., 2012). Although the engineered plants' agronomical performances are challenged, enhanced cell wall deconstruction have been found in down-regulation of CCR and CCoAOMT in maize (Park et al., 2012; Li et al., 2013), CAD, 4CL and COMT in switchgrass (Fu et al., 2011a,b; Xu et al., 2011), HCT in *Arabidopsis* (Gallego-Giraldo et al., 2011), C3'H and C4H in *Eucalyptus* (Sykes et al., 2015). In addition, inclusion of new lignin precursors has also been used to facilitate lignin depolymerization thereby reduce biomass recalcitrance. With incorporation of several flavonoids and gallate derivatives with easily cleavable ester bonds, the artificially lignified cell wall had enhanced delignification, ruminal and/or enzymatic digestibility and fermentability (Grabber et al., 2010; Elumalai et al., 2012).

## SUMMARY AND PERSPECTIVES

Although the study of lignin's influence on cell wall digestibility has been advanced recently, the correlation of lignin structure and biomass recalcitrance remain complicated due to lignin structural variety and cell wall complexity. According to these studies, the physicochemical properties of lignin are strongly related with recalcitrance (Figure 2). It should be noted that this figure only represents the major findings in certain case and it cannot be solely used as a metric to evaluate biomass recalcitrance as these factors and other unknown factors usually contribute together to cell wall recalcitrance. The impact of

lignin on biomass recalcitrance could be very different depending upon plant species and pretreatment techniques. To better understand the cause-effect relation between one property of lignin and biomass recalcitrance, it is desirable to minimize the side effects from other cell wall properties and lignin properties.

## AUTHOR CONTRIBUTIONS

AR conceived the review and reviewed it for technical merit. ML and YP shared in writing the article.

## REFERENCES

- Azarpira, A., Lu, F., and Ralph, J. (2011). Reactions of dehydrodiferulates with ammonia. *Org. Biomol. Chem.* 9, 6779–6787. doi: 10.1039/c1ob05677h
- Barsberg, S., Selig, M. J., and Felby, C. (2013). Impact of lignins isolated from pretreated lignocelluloses on enzymatic cellulose saccharification. *Biotechnol. Lett.* 35, 189–195. doi: 10.1007/s10529-012-1061-x
- Berlin, A., Balakshin, M., Gilkes, N., Kadla, J., Maximenko, V., Kubo, S., et al. (2006). Inhibition of cellulase, xylanase and  $\beta$ -glucosidase activities by softwood lignin preparations. *J. Biotechnol.* 125, 198–209. doi: 10.1016/j.jbiotec.2006.02.021
- Besombes, S., and Mazeau, K. (2005a). The cellulose/lignin assembly assessed by molecular modeling. Part 1: adsorption of a threo guaiacyl  $\beta$ -O-4 dimer onto a  $\beta$  cellulose whisker. *Plant Physiol. Biochem.* 43, 299–308. doi: 10.1016/j.plaphy.2005.02.005
- Besombes, S., and Mazeau, K. (2005b). The cellulose/lignin assembly assessed by molecular modeling. Part 2: seeking for evidence of organization of lignin molecules at the interface with cellulose. *Plant Physiol. Biochem.* 43, 277–286. doi: 10.1016/j.plaphy.2005.02.004
- Boerjan, W., Ralph, J., and Baucher, M. (2003). Lignin biosynthesis. *Annu. Rev. Plant Biol.* 54, 519–546. doi: 10.1146/annurev-arplant.54.031902.134938
- Bonawit, N. D., and Chapple, C. (2010). The genetics of lignin biosynthesis: connecting genotype to phenotype. *Annu. Rev. Genet.* 44, 337–363. doi: 10.1146/annurev-genet-102209-163508
- Buranov, A. U., and Mazza, G. (2008). Lignin in straw of herbaceous crops. *Ind. Crops Prod.* 28, 237–259. doi: 10.1016/j.indcrop.2008.03.008
- Cesarino, I., Araújo, P., Domingues Júnior, A. P., and Mazzafera, P. (2012). An overview of lignin metabolism and its effect on biomass recalcitrance. *Braz. J. Bot.* 35, 303–311. doi: 10.1590/S0100-84042012000400003
- Chundawat, S. P., Donohoe, B. S., da Costa Sousa, L., Elder, T., Agarwal, U. P., Lu, F., et al. (2011). Multi-scale visualization and characterization of lignocellulosic plant cell wall deconstruction during thermochemical pretreatment. *Energy Environ. Sci.* 4, 973–984. doi: 10.1039/c0ee00574f
- de O Buanafina M. M. (2009). Feruloylation in grasses: current and future perspectives. *Mol. Plant* 2, 861–872. doi: 10.1093/mp/ssp067
- Donohoe, B. S., Decker, S. R., Tucker, M. P., Himmel, M. E., and Vinzant, T. B. (2008). Visualizing lignin coalescence and migration through maize cell walls following thermochemical pretreatment. *Biotechnol. Bioeng.* 101, 913–925. doi: 10.1002/bit.21959
- Elumalai, S., Tobimatsu, Y., Grabber, J. H., Pan, X., and Ralph, J. (2012). Epigallocatechin gallate incorporation into lignin enhances the alkaline delignification and enzymatic saccharification of cell walls. *Biotechnol. Biofuels* 5:59. doi: 10.1186/1754-6834-5-59
- Fu, C., Mielenz, J. R., Xiao, X., Ge, Y., Hamilton, C. Y., Rodriguez, M., et al. (2011a). Genetic manipulation of lignin reduces recalcitrance and improves ethanol production from switchgrass. *Proc. Natl. Acad. Sci. U.S.A.* 108, 3803–3808. doi: 10.1073/pnas.1100310108
- Fu, C., Xiao, X., Xi, Y., Ge, Y., Chen, F., Bouton, J., et al. (2011b). Downregulation of cinnamyl alcohol dehydrogenase (CAD) leads to improved saccharification efficiency in switchgrass. *Bioenerg. Res.* 4, 153–164. doi: 10.1007/s12155-010-9109-z
- Gallego-Giraldo, L., Escamilla-Trevino, L., Jackson, L. A., and Dixon, R. A. (2011). Salicylic acid mediates the reduced growth of lignin down-regulated plants. *Proc. Natl. Acad. Sci. U.S.A.* 108, 20814–20819. doi: 10.1073/pnas.1117873108
- Grabber, J. H., Schatz, P. F., Kim, H., Lu, F., and Ralph, J. (2010). Identifying new lignin bioengineering targets: 1. Monolignol-substitute impacts on lignin formation and cell wall fermentability. *BMC Plant Biol.* 10:114. doi: 10.1186/1471-2229-10-114
- Guo, F., Shi, W., Sun, W., Li, X., Wang, F., Zhao, J., et al. (2014). Differences in the adsorption of enzymes onto lignins from diverse types of lignocellulosic biomass and the underlying mechanism. *Biotechnol. Biofuels* 7:38. doi: 10.1186/1754-6834-7-38
- Güven, G., Prodanovic, R., and Schwaneberg, U. (2010). Protein engineering—an option for enzymatic biofuel cell design. *Electroanalysis* 22, 765–775. doi: 10.1002/elan.200980017
- Himmel, M. E., Ding, S.-Y., Johnson, D. K., Adney, W. S., Nimlos, M. R., Brady, J. W., et al. (2007). Biomass recalcitrance: engineering plants and enzymes for biofuels production. *Science* 315, 804–807. doi: 10.1126/science.1137016
- Hu, F., Jung, S., and Ragauskas, A. (2012). Pseudo-lignin formation and its impact on enzymatic hydrolysis. *Bioresour. Technol.* 117, 7–12. doi: 10.1016/j.biortech.2012.04.037
- Huang, C., He, J., Min, D., Lai, C., and Yong, Q. (2016). Understanding the nonproductive enzyme adsorption and physicochemical properties of residual lignins in moso bamboo pretreated with sulfuric acid and kraft pulping. *Appl. Biochem. Biotechnol.* doi: 10.1007/s12010-016-2183-8. [Epub ahead of print].
- Jiang, B., Wang, W., Gu, F., Cao, T., and Jin, Y. (2016). Comparison of the substrate enzymatic digestibility and lignin structure of wheat straw stems and leaves pretreated by green liquor. *Bioresour. Technol.* 199, 181–187. doi: 10.1016/j.biortech.2015.08.104
- Jung, H., Mertens, D., and Phillips, R. (2011). Effect of reduced ferulate-mediated lignin/arabinoxylan cross-linking in corn silage on feed intake, digestibility, and milk production. *J. Dairy Sci.* 94, 5124–5137. doi: 10.3168/jds.2011-4495
- Jung, H., and Phillips, R. (2010). Putative seedling ferulate ester (*sfe*) maize mutant: morphology, biomass yield, and stover cell wall composition and rumen degradability. *Crop Sci.* 50, 403–418. doi: 10.2135/cropsci2009.04.0191
- Kim, T. H. (2012). Comparison of inhibition effects of various isolated lignins on enzymatic hydrolysis of cellulose. *Korean J. Chem. Eng.* 29, 82–88. doi: 10.1007/s11814-011-0150-1
- Ko, J. K., Kim, Y., Ximenes, E., and Ladisch, M. R. (2015). Effect of liquid hot water pretreatment severity on properties of hardwood lignin and enzymatic hydrolysis of cellulose. *Biotechnol. Bioeng.* 112, 252–262. doi: 10.1002/bit.25349
- Kumar, L., Arantes, V., Chandra, R., and Saddler, J. (2012). The lignin present in steam pretreated softwood binds enzymes and limits cellulose accessibility. *Bioresour. Technol.* 103, 201–208. doi: 10.1016/j.biortech.2011.09.091
- Lai, C., Tu, M., Li, M., and Yu, S. (2014). Remarkable solvent and extractable lignin effects on enzymatic digestibility of organosolv pretreated hardwood. *Bioresour. Technol.* 156, 92–99. doi: 10.1016/j.biortech.2014.01.030
- Langan, P., Petridis, L., O'Neill, H. M., Pingali, S. V., Foston, M., Nishiyama, Y., et al. (2014). Common processes drive the thermochemical pretreatment of lignocellulosic biomass. *Green Chem.* 16, 63–68. doi: 10.1039/C3GC41962B
- Li, H., Pu, Y., Kumar, R., Ragauskas, A. J., and Wyman, C. E. (2014). Investigation of lignin deposition on cellulose during hydrothermal pretreatment, its effect

## ACKNOWLEDGMENTS

We thank Dr. Xianzhi Meng and Dr. Chang Geun Yoo for assistance on chemical structures drawing. This study was supported and performed as part of the BioEnergy Science Center (BESC). The BESC is a U.S. Department of Energy Bioenergy Research Center supported by the Office of Biological and Environmental Research in the DOE Office of Science. Oak Ridge National Laboratory is managed by UT-Battelle, LLC under contract No. DE-AC05-00OR22725 with the U.S. Department of Energy.



- on cellulose hydrolysis, and underlying mechanisms. *Biotechnol. Bioeng.* 111, 485–492. doi: 10.1002/bit.25108
- Li, M., Foster, C., Kelkar, S., Pu, Y., Holmes, D., Ragauskas, A., et al. (2012). Structural characterization of alkaline hydrogen peroxide pretreated grasses exhibiting diverse lignin phenotypes. *Biotechnol. Biofuels* 5:38. doi: 10.1186/1754-6834-5-38
- Li, M., Pu, Y., Yoo, C. G., and Ragauskas, A. J. (2016). The occurrence of tricin and its derivatives in plants. *Green Chem.* 18, 1439–1454. doi: 10.1039/C5GC03062E
- Li, M., Si, S., Hao, B., Zha, Y., Wan, C., Hong, S., et al. (2014). Mild alkali-pretreatment effectively extracts guaiacyl-rich lignin for high lignocellulose digestibility coupled with largely diminishing yeast fermentation inhibitors in *Miscanthus*. *Bioresour. Technol.* 169, 447–454. doi: 10.1016/j.biortech.2014.07.017
- Li, X., Chen, W., Zhao, Y., Xiang, Y., Jiang, H., Zhu, S., et al. (2013). Downregulation of caffeoyl-CoA O-methyltransferase (CCoAOMT) by RNA interference leads to reduced lignin production in maize straw. *Genet. Mol. Biol.* 36, 540–546. doi: 10.1590/S1415-47572013005000039
- Li, X., Ximenes, E., Kim, Y., Slininger, M., Meilan, R., Ladisch, M., et al. (2010). Lignin monomer composition affects *Arabidopsis* cell-wall degradability after liquid hot water pretreatment. *Biotechnol. Biofuels* 3:27. doi: 10.1186/1754-6834-3-27
- Li, Y., Qi, B., Luo, J., and Wan, Y. (2016). Effect of alkali lignins with different molecular weights from alkali pretreated rice straw hydrolyzate on enzymatic hydrolysis. *Bioresour. Technol.* 200, 272–278. doi: 10.1016/j.biortech.2015.10.038
- Liu, H., Zhu, J., and Fu, S. (2010). Effects of lignin–metal complexation on enzymatic hydrolysis of cellulose. *J. Agric. Food Chem.* 58, 7233–7238. doi: 10.1021/jf1001588
- Mansfield, S. D., Kang, K. Y., and Chapple, C. (2012). Designed for deconstruction–poplar trees altered in cell wall lignification improve the efficacy of bioethanol production. *New Phytol.* 194, 91–101. doi: 10.1111/j.1469-8137.2011.04031.x
- Martínez, P. M., Punt, A. M., Kabel, M. A., and Gruppen, H. (2016). Deconstruction of lignin linked p-coumarates, ferulates and xylan by NaOH enhances the enzymatic conversion of glucan. *Bioresour. Technol.* 216, 44–51. doi: 10.1016/j.biortech.2016.05.040
- McCann, M. C., and Carpita, N. C. (2015). Biomass recalcitrance: a multi-scale, multi-factor and conversion-specific property. *J. Exp. Bot.* 66, 4109–4118. doi: 10.1093/jxb/erv267
- Min, D., Yang, C., Shi, R., Jameel, H., Chiang, V., and Chang, H. (2013). The elucidation of the lignin structure effect on the cellulase-mediated saccharification by genetic engineering poplars (*Populus nigra* L. × *Populus maximowiczii* A.). *Biomass Bioenerg.* 58, 52–57. doi: 10.1016/j.biombioe.2013.08.019
- Mood, S. H., Golfeshan, A. H., Tabatabaei, M., Jouzani, G. S., Najafi, G. H., Gholami, M., et al. (2013). Lignocellulosic biomass to bioethanol, a comprehensive review with a focus on pretreatment. *Renew. Sust. Energ. Rev.* 27, 77–93. doi: 10.1016/j.rser.2013.06.033
- Mosier, N., Wyman, C., Dale, B., Elander, R., Lee, Y., Holtzapple, M., et al. (2005). Features of promising technologies for pretreatment of lignocellulosic biomass. *Bioresour. Technol.* 96, 673–686. doi: 10.1016/j.biortech.2004.06.025
- Moxley, G., Gaspar, A. R., Higgins, D., and Xu, H. (2012). Structural changes of corn stover lignin during acid pretreatment. *J. Ind. Microbiol. Biotechnol.* 39, 1289–1299. doi: 10.1007/s10295-012-1131-z
- Nakagame, S., Chandra, R. P., Kadla, J. F., and Saddler, J. N. (2011b). Enhancing the enzymatic hydrolysis of lignocellulosic biomass by increasing the carboxylic acid content of the associated lignin. *Biotechnol. Bioeng.* 108, 538–548. doi: 10.1002/bit.22981
- Nakagame, S., Chandra, R. P., Kadla, J. F., and Saddler, J. N. (2011c). The isolation, characterization and effect of lignin isolated from steam pretreated Douglas-fir on the enzymatic hydrolysis of cellulose. *Bioresour. Technol.* 102, 4507–4517. doi: 10.1016/j.biortech.2010.12.082
- Nakagame, S., Chandra, R. P., and Saddler, J. N. (2010). The effect of isolated lignins, obtained from a range of pretreated lignocellulosic substrates, on enzymatic hydrolysis. *Biotechnol. Bioeng.* 105, 871–879. doi: 10.1002/bit.22626
- Nakagame, S., Chandra, R., Saddler, J., Zhu, J., Zhang, X., and Pan, X. (2011a). “The influence of lignin on the enzymatic hydrolysis of pretreated biomass substrates,” in *Sustainable Production of Fuels, Chemicals, and Fibers from Forest Biomass*, ACS Symposium Series, eds J. Y. Zhu, X. Zhang, and X. Pan (Washington, DC: Oxford University Press), 145–167. doi: 10.1021/bk-2011-1067.ch006
- Papa, G., Varanasi, P., Sun, L., Cheng, G., Stavila, V., Holmes, B., et al. (2012). Exploring the effect of different plant lignin content and composition on ionic liquid pretreatment efficiency and enzymatic saccharification of *Eucalyptus globulus* L. mutants. *Bioresour. Technol.* 117, 352–359. doi: 10.1016/j.biortech.2012.04.065
- Pareek, N., Gillgren, T., and Jönsson, L. J. (2013). Adsorption of proteins involved in hydrolysis of lignocellulose on lignins and hemicelluloses. *Bioresour. Technol.* 148, 70–77. doi: 10.1016/j.biortech.2013.08.121
- Park, S.-H., Mei, C., Pauly, M., Ong, R. G., Dale, B. E., Sabzikar, R., et al. (2012). Downregulation of maize cinnamoyl-coenzyme A reductase via RNA interference technology causes brown midrib and improves ammonia fiber expansion-pretreated conversion into fermentable sugars for biofuels. *Crop Sci.* 52, 2687–2701. doi: 10.2135/cropsci2012.04.0253
- Petrik, D. L., Karlen, S. D., Cass, C. L., Padmakshan, D., Lu, F., Liu, S., et al. (2014). p-Coumaroyl-CoA: monolignol transferase (PMT) acts specifically in the lignin biosynthetic pathway in *Brachypodium distachyon*. *Plant J.* 77, 713–726. doi: 10.1111/tpj.12420
- Phitsuwan, P., Sakka, K., and Ratanakhanokchai, K. (2013). Improvement of lignocellulosic biomass in planta: a review of feedstocks, biomass recalcitrance, and strategic manipulation of ideal plants designed for ethanol production and processability. *Biomass Bioenerg.* 58, 390–405. doi: 10.1016/j.biombioe.2013.08.027
- Pingali, S. V., Urban, V. S., Heller, W. T., McGaughey, J., O'Neill, H., Foston, M., et al. (2010). Breakdown of cell wall nanostructure in dilute acid pretreated biomass. *Biomacromolecules* 11, 2329–2335. doi: 10.1021/bm100455h
- Pu, Y., Hu, F., Huang, F., Davison, B. H., and Ragauskas, A. J. (2013). Assessing the molecular structure basis for biomass recalcitrance during dilute acid and hydrothermal pretreatments. *Biotechnol. Biofuels* 6:15. doi: 10.1186/1754-6834-6-15
- Pu, Y., Hu, F., Huang, F., and Ragauskas, A. J. (2015). Lignin structural alterations in thermochemical pretreatments with limited delignification. *Bioenerg. Res.* 8, 992–1003. doi: 10.1007/s12155-015-9655-5
- Pu, Y., Kosa, M., Kalluri, U. C., Tuskan, G. A., and Ragauskas, A. J. (2011). Challenges of the utilization of wood polymers: how can they be overcome? *Appl. Microbiol. Biotechnol.* 91, 1525–1536. doi: 10.1007/s00253-011-3350-z
- Qin, L., Li, W.-C., Zhu, J.-Q., Liang, J.-N., Li, B.-Z., and Yuan, Y.-J. (2015). Ethylenediamine pretreatment changes cellulose allomorph and lignin structure of lignocellulose at ambient pressure. *Biotechnol. Biofuels* 8, 174. doi: 10.1186/s13068-015-0359-z
- Ragauskas, A. J., Williams, C. K., Davison, B. H., Britovsek, G., Cairney, J., Eckert, C. A., et al. (2006). The path forward for biofuels and biomaterials. *Science* 311, 484–489. doi: 10.1126/science.1114736
- Rahikainen, J. L., Martin-Sampedro, R., Heikkinen, H., Rovio, S., Marjamaa, K., Tamminen, T., et al. (2013). Inhibitory effect of lignin during cellulose bioconversion: the effect of lignin chemistry on non-productive enzyme adsorption. *Bioresour. Technol.* 133, 270–278. doi: 10.1016/j.biortech.2013.01.075
- Ralph, J. (2010). Hydroxycinnamates in lignification. *Phytochem. Rev.* 9, 65–83. doi: 10.1007/s11101-009-9141-9
- Samuel, R., Cao, S., Das, B. K., Hu, F., Pu, Y., and Ragauskas, A. J. (2013). Investigation of the fate of poplar lignin during autohydrolysis pretreatment to understand the biomass recalcitrance. *RSC Adv.* 3, 5305–5309. doi: 10.1039/c3ra40578h
- Samuel, R., Pu, Y., Raman, B., and Ragauskas, A. J. (2010). Structural characterization and comparison of switchgrass ball-milled lignin before and after dilute acid pretreatment. *Appl. Biochem. Biotechnol.* 162, 62–74. doi: 10.1007/s12010-009-8749-y
- Sannigrahi, P., Kim, D. H., Jung, S., and Ragauskas, A. (2011). Pseudo-lignin and pretreatment chemistry. *Energy Environ. Sci.* 4, 1306–1310. doi: 10.1039/C0EE00378F
- Santos, R. B., Capanema, E. A., Balakshin, M. Y., Chang, H.-M., and Jameel, H. (2011). Effect of hardwoods characteristics on kraft pulping process: emphasis on lignin structure. *BioResources* 6, 3623–3637. doi: 10.1021/bm100455h
- Santos, R. B., Lee, J. M., Jameel, H., Chang, H.-M., and Lucia, L. A. (2012). Effects of hardwood structural and chemical characteristics on enzymatic

- hydrolysis for biofuel production. *Bioresour. Technol.* 110, 232–238. doi: 10.1016/j.biortech.2012.01.085
- Shi, J., Pattathil, S., Ramakrishnan, P., Anderson, N., Im Kim, J., Venkatachalam, S., et al. (2016). Impact of engineered lignin composition on biomass recalcitrance and ionic liquid pretreatment efficiency. *Green Chem.* 18, 4884–4895. doi: 10.1039/C6GC01193D
- Simmons, B. A., Loque, D., and Ralph, J. (2010). Advances in modifying lignin for enhanced biofuel production. *Curr. Opin. Plant Biol.* 13, 312–319. doi: 10.1016/j.pbi.2010.03.001
- Studer, M. H., DeMartini, J. D., Davis, M. F., Sykes, R. W., Davison, B., Keller, M., et al. (2011). Lignin content in natural *Populus* variants affects sugar release. *Proc. Natl. Acad. Sci. U.S.A.* 108, 6300–6305. doi: 10.1073/pnas.1009252108
- Sykes, R. W., Gjersing, E. L., Foutz, K., Rottmann, W. H., Kuhn, S. A., Foster, C. E., et al. (2015). Down-regulation of p-coumaroyl quinate/shikimate 3'-hydroxylase (C3' H) and cinnamate 4-hydroxylase (C4H) genes in the lignin biosynthetic pathway of *Eucalyptus urophylla* × *E. grandis* leads to improved sugar release. *Biotechnol. Biofuels* 8, 128. doi: 10.1186/s13068-015-0316-x
- Taboada, A., Novo-Uzal, E., Flores, G., Loureda, M., Ros Barceló, A., Masa, A., et al. (2010). Digestibility of silages in relation to their hydroxycinnamic acid content and lignin composition. *J. Sci. Food Agric.* 90, 1155–1162. doi: 10.1002/jsfa.3933
- Tobimatsu, Y., Elumalai, S., Grabber, J. H., Davidson, C. L., Pan, X., and Ralph, J. (2012). Hydroxycinnamate conjugates as potential monolignol replacements: *in vitro* lignification and cell wall studies with rosmarinic acid. *ChemSusChem* 5, 676–686. doi: 10.1002/cssc.201100573
- Trajano, H. L., Engle, N. L., Foston, M., Ragauskas, A. J., Tschaplinski, T. J., and Wyman, C. E. (2013). The fate of lignin during hydrothermal pretreatment. *Biotechnol. Biofuels* 6:110. doi: 10.1186/1754-6834-6-110
- Vanholme, R., Demedts, B., Morreel, K., Ralph, J., and Boerjan, W. (2010). Lignin biosynthesis and structure. *Plant Physiol.* 153, 895–905. doi: 10.1104/pp.110.155119
- Wang, W., Zhu, Y., Du, J., Yang, Y., and Jin, Y. (2015). Influence of lignin addition on the enzymatic digestibility of pretreated lignocellulosic biomasses. *Bioresour. Technol.* 181, 7–12. doi: 10.1016/j.biortech.2015.01.026
- Wang, Z., Zhu, J., Fu, Y., Qin, M., Shao, Z., Jiang, J., et al. (2013). Lignosulfonate-mediated cellulase adsorption: enhanced enzymatic saccharification of lignocellulose through weakening nonproductive binding to lignin. *Biotechnol. Biofuels* 6:156. doi: 10.1186/1754-6834-6-156
- Wilkerson, C., Mansfield, S., Lu, F., Withers, S., Park, J.-Y., Karlen, S., et al. (2014). Monolignol ferulate transferase introduces chemically labile linkages into the lignin backbone. *Science* 344, 90–93. doi: 10.1126/science.1250161
- Wu, Z., Zhang, M., Wang, L., Tu, Y., Zhang, J., Xie, G., et al. (2013). Biomass digestibility is predominantly affected by three factors of wall polymer features distinctive in wheat accessions and rice mutants. *Biotechnol. Biofuels* 6:183. doi: 10.1186/1754-6834-6-183
- Xu, B., Escamilla-Treviño, L. L., Sathitsuksanoh, N., Shen, Z., Shen, H., Percival Zhang, Y. H., et al. (2011). Silencing of 4-coumarate: coenzyme A ligase in switchgrass leads to reduced lignin content and improved fermentable sugar yields for biofuel production. *New Phytol.* 192, 611–625. doi: 10.1111/j.1469-8137.2011.03830.x
- Xu, N., Zhang, W., Ren, S., Liu, F., Zhao, C., Liao, H., et al. (2012). Hemicelluloses negatively affect lignocellulose crystallinity for high biomass digestibility under NaOH and H<sub>2</sub>SO<sub>4</sub> pretreatments in *Miscanthus*. *Biotechnol. Biofuels* 5:58. doi: 10.1186/1754-6834-5-58
- Yang, Q., and Pan, X. (2016). Correlation between lignin physicochemical properties and inhibition to enzymatic hydrolysis of cellulose. *Biotechnol. Bioeng.* 113, 1213–1224. doi: 10.1002/bit.25903
- Yeh, T.-F., Chang, M.-J., and Chang, W.-J. (2014). Comparison of dilute acid and sulfite pretreatments on acacia confusa for biofuel application and the influence of its extractives. *J. Agric. Food Chem.* 62, 10768–10775. doi: 10.1021/jf504461c
- Yoo, C. G., Kim, H., Lu, F., Azarpira, A., Pan, X., Oh, K. K., et al. (2016a). Understanding the physicochemical characteristics and the improved enzymatic saccharification of corn stover pretreated with aqueous and gaseous ammonia. *Bioenerg. Res.* 9, 67–76. doi: 10.1007/s12155-015-9662-6
- Yoo, C. G., Pu, Y., Li, M., and Ragauskas, A. J. (2016b). Elucidating structural characteristics of biomass using solution-state 2 D Nmr with a mixture of deuterated dimethylsulfoxide and hexamethylphosphoramide. *ChemSusChem* 9, 1090–1095. doi: 10.1002/cssc.201600135
- Yu, Z., Gwak, K.-S., Treasure, T., Jameel, H., Chang, H.-m., and Park, S. (2014). Effect of lignin chemistry on the enzymatic hydrolysis of woody biomass. *ChemSusChem* 7, 1942–1950. doi: 10.1002/cssc.201400042
- Zeng, Y., Zhao, S., Yang, S., and Ding, S.-Y. (2014). Lignin plays a negative role in the biochemical process for producing lignocellulosic biofuels. *Curr. Opin. Biotechnol.* 27, 38–45. doi: 10.1016/j.copbio.2013.09.008
- Zhang, Y., Culhaoglu, T., Pollet, B., Melin, C., Denoue, D., Barriere, Y., et al. (2011). Impact of lignin structure and cell wall reticulation on maize cell wall degradability. *J. Agric. Food Chem.* 59, 10129–10135. doi: 10.1021/jf2028279
- Zhou, H., Lou, H., Yang, D., Zhu, J., and Qiu, X. (2013). Lignosulfonate to enhance enzymatic saccharification of lignocelluloses: role of molecular weight and substrate lignin. *Ind. Eng. Chem. Res.* 52, 8464–8470. doi: 10.1021/ie401085k
- Ziebell, A., Gracom, K., Katahira, R., Chen, F., Pu, Y., Ragauskas, A., et al. (2010). Increase in 4-coumaryl alcohol units during lignification in alfalfa (*Medicago sativa*) alters the extractability and molecular weight of lignin. *J. Biol. Chem.* 285, 38961–38968. doi: 10.1074/jbc.M110.137315

**Disclosure:** This manuscript has been authored by UT-Battelle, LLC under contract no. DE-AC05-00OR22725 with the U.S. Department of Energy. The publisher, by accepting the article for publication, acknowledges that the United States Government retains a non-exclusive, paid-up, irrevocable, world-wide license to publish or reproduce the published form of this manuscript, or allow others to do so, for United States Government purposes. The Department of Energy will provide public access to these results of federally sponsored research in Accordance with the DOE Public Access Plan (<http://energy.gov/downloads/doe-public-access-plan>).

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2016 Li, Pu and Ragauskas. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.